



TOTAL SYNTHESIS OF LIPOPHILIC MASKED NAADP

DERIVATIVES

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Abstract

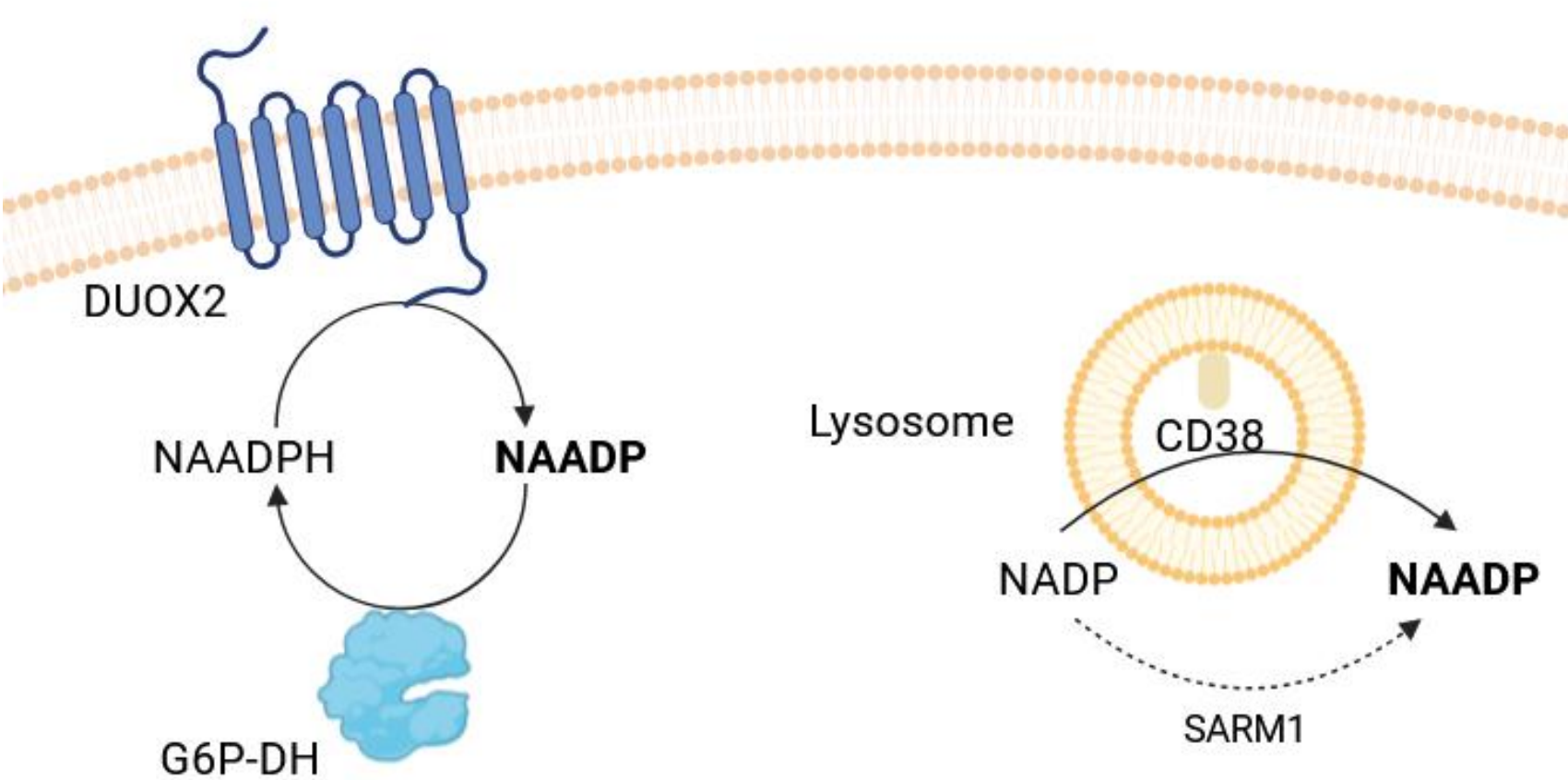
Nicotinic acid adenine dinucleotide phosphate (NAADP) is the most potent Ca^{2+} -releasing second messenger.^[1] The release of Ca^{2+} by NAADP was verified in different eukaryotic cell types, for example in human T-lymphocytes.^[2] To study its cellular effects, NAADP must be present in the cells, which is challenging due to its polarity. In our group, a new **masked compound** was developed:

Membrane permeable STabilized bio-rEversible pRotected NAADP (MASTER-NAADP)^[3]

The MASTER concept introduced several modifications leading to a **higher stability** compared to previously applied masking concept, e.g. the AM-approach.^[3]

Biosynthesis pathways of NAADP^[4]

Proposed biosynthesis pathways of NAADP e.g. after binding of an antigen presenting cell to a T-cell

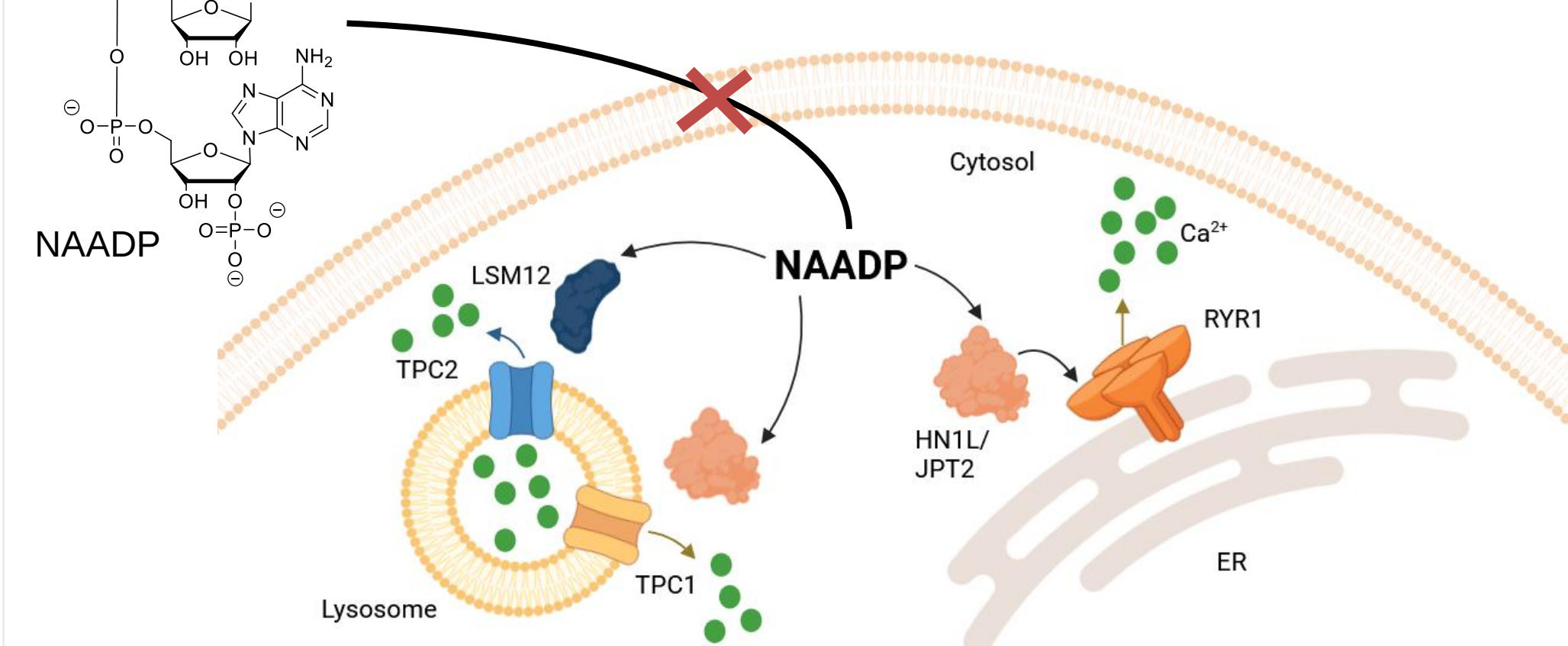


DUOX2: dual oxidase 2
NAADP: Nicotinic acid adenine dinucleotide phosphate
NADP: Nicotinamide adenine dinucleotide phosphate
G6P-DH: glucose 6-phosphate dehydrogenase

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NAADP induced Ca^{2+} -release^[4]

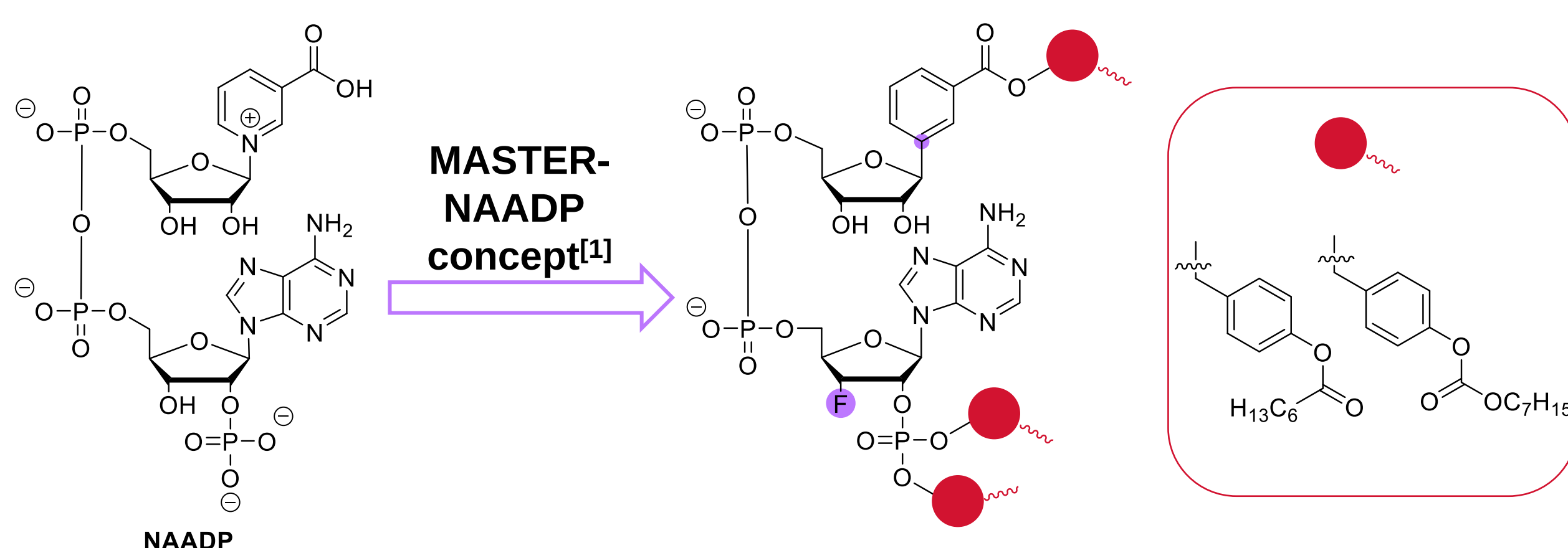
NAADP cannot passively diffuse through the cell membrane → **membrane-permeable NAADP**



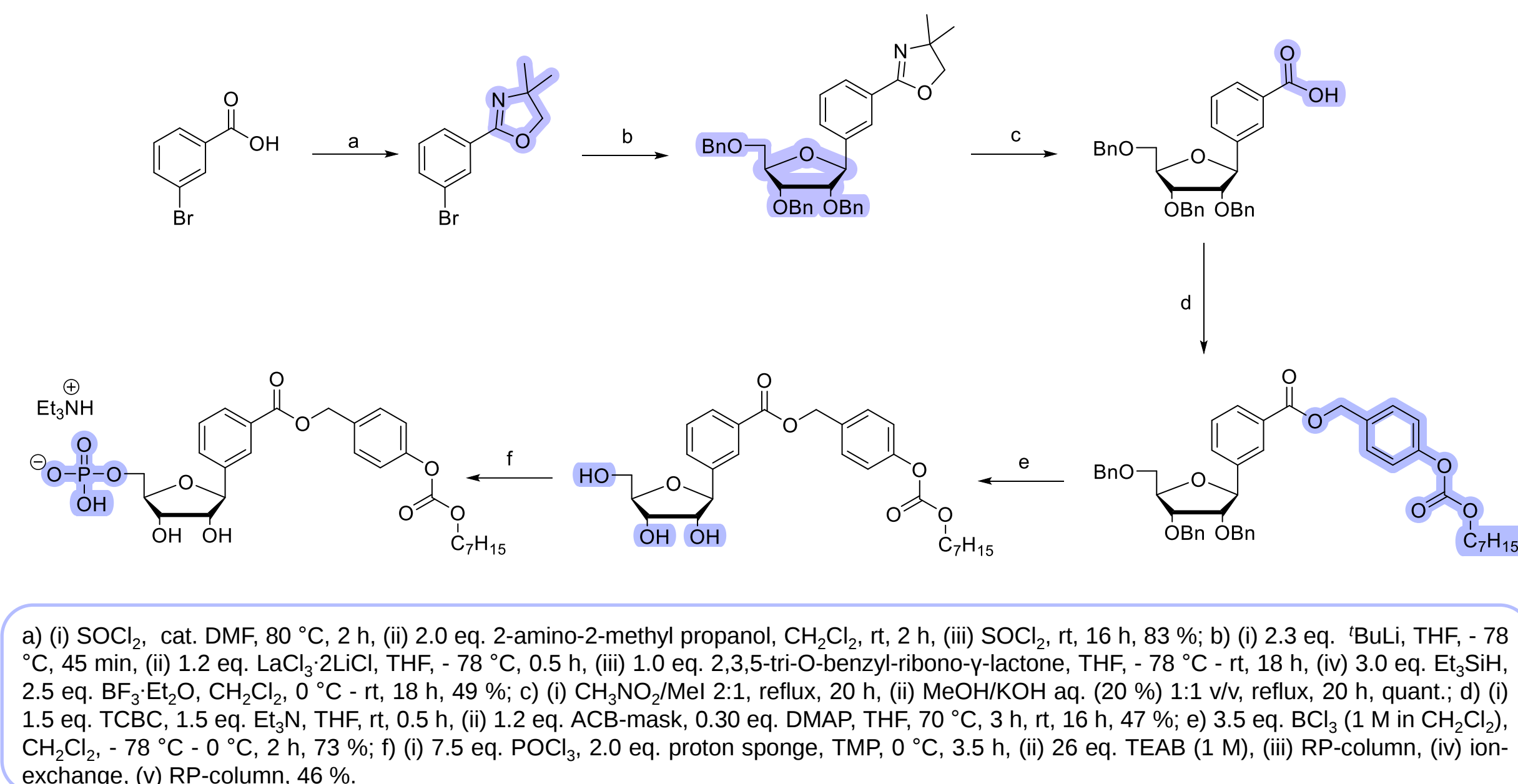
LSM12: Sm-like 12
TPC2: two-pore channel 2
HN1L/JPT2: 1-like/Jupiter microtubule-associated homolog 2
RyR1: Ryanodine receptor 1

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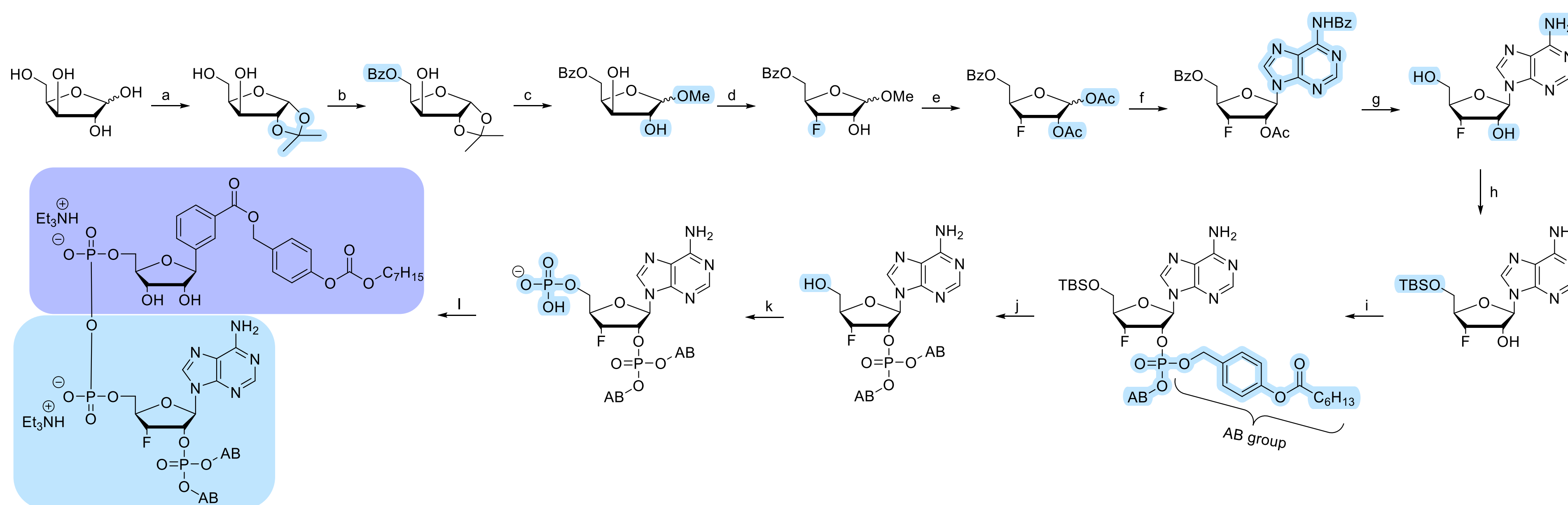
Objective



Synthesis “Northern” building block^[3,5]

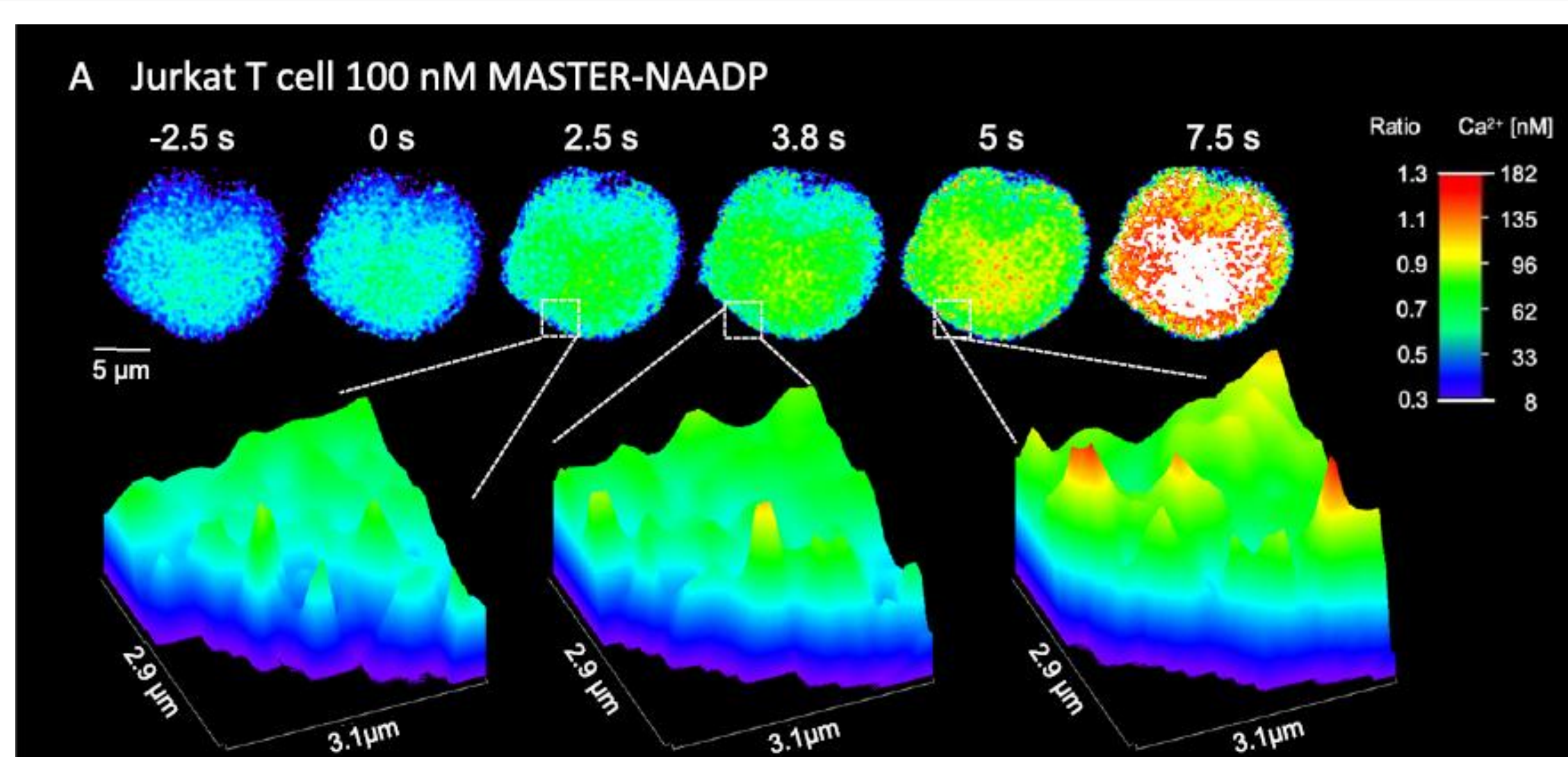


Synthesis “Southern” building block and coupling of the NAADP derivative^[3,5]

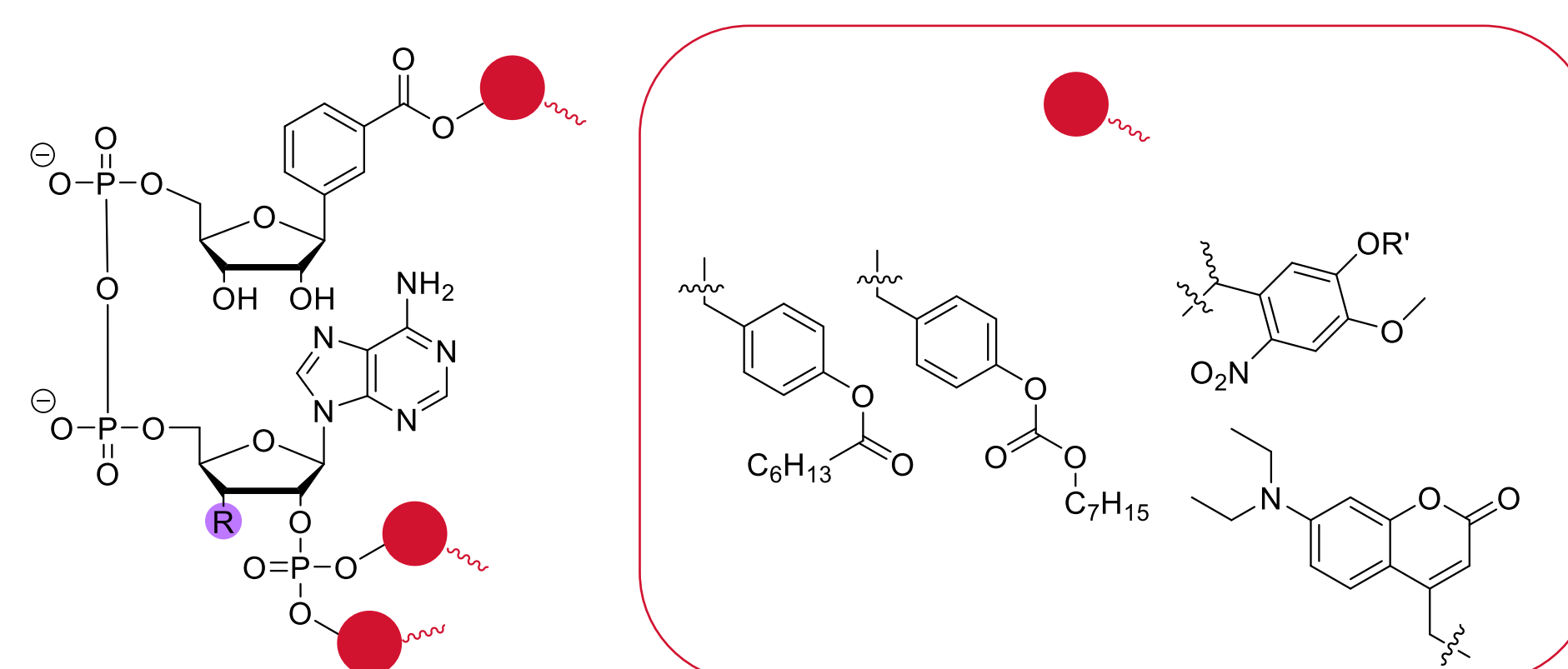


a) (i) 0.10 eq. CuSO_4 , 2.8 eq. conc. H_2SO_4 , acetone, 3h, rt, (ii) HCl-solution (0.2%), 6h, rt, 99%; b) 1.5 eq. Et_3N , 1.1 eq. BzCl , CH_2Cl_2 , 1 h, 0 °C, 92 %; c) 0.2 eq. I_2 , MeOH, 4 h, 70 °C, 86 %; d) 1.5 eq. DAST, dry CH_2Cl_2 , 20h, -78 °C - rt, 43 %; e) 4.7 eq. Ac_2O , 4.7 eq. conc. H_2SO_4 , AcOH, 19 h, rt, 93 %; f) (i) 1.5 eq. *N*-benzoyladenine, 1.2 eq. BSA, $\text{C}_2\text{H}_5\text{Cl}$, 1 h, 90 °C, (ii) 1.0 eq. ribose derivative, 4.0 eq. TMSOTf, $\text{C}_2\text{H}_5\text{Cl}$, 5.5 h, 90 °C, 61 %; g) ammonia 7 N in MeOH, 28 h, 70 °C, 84%; h) 1.5 eq. TBSCl, dry pyridine, 24 h, rt, 71 %; i) (i) 1.3 eq. amidite, 1.2 eq. pyridinium trifluoroacetate, CH_2Cl_2 , 50 min, rt, (ii) 1.5 eq. tert-butylhydroperoxide (5.0M in decane), 0 °C, 1 h, 87 %; j) 5.2 eq. triethylamine-trihydrofluoride, CH_2Cl_2 , 19h, rt, 81 %; k) 4.0 eq. POCl_3 , TMP, 6.5 h, 0 °C, 31 %; l) (i) 16 eq. Et_3N , 20 eq. TFAA, CH_3CN , 0 °C, 15 min, (ii) 1.0 eq. “northern” monophosphate, CH_3CN , 0 °C, 15 min, (iii) 6 eq. NMI, 10 eq. Et_3N , CH_3CN , rt, 15 min, (iv) 0.9 eq. diphosphate, $\text{CH}_3\text{CN}/\text{DMF}$ 5:1, rt, 3 h, 57 %.

Ca^{2+} -release in Jurkat T cells^[3]



Outlook



References

[1] R. Aarhus, R. M. Graeff, D. M. Dickey, T. F. Walseth, C. L. Hon, *Journal of Biological Chemistry* **1995**, 270, 30327; [2] A. Gasser, S. Bruhn, A. H. Guse, *Journal of Biological Chemistry* **2006**, 281, 16906; [3] S. Krukenberg, F. Möckl, M. Weiß, P. Dekiert, M. Hofmann, F. Gerlach, K. J. Winterberg, D. Kovacevic, I. Khansahib, B. Troost et al., *Nat Commun* **2024**, 15, 8008; [4] A. H. Guse, *Cells* **2022**, 11; [5] M. Yoshikawa, T. Kato, T. Takenishi, *Tetrahedron Letters* **1967**, 8, 5065.